
Alzheimer's Disease: Etiology, Neuropathology and Pathogenesis

Olivia Sheppard • Michael Coleman

John van Geest Centre for Brain Repair, Department of Clinical Neurosciences, University of Cambridge, Cambridge, UK

Author for correspondence: Olivia Sheppard, John van Geest Centre for Brain Repair, Department of Clinical Neurosciences, University of Cambridge, Cambridge, UK.
Email: os356@cam.ac.uk

Doi: <https://doi.org/10.36255/exonpublications.alzheimersdisease.2020.ch1>

Abstract: Alzheimer's disease is the most common form of dementia and the most common neurodegenerative disease. It manifests as a decline in short-term memory and cognition that impairs daily behavior. Most cases of Alzheimer's disease are sporadic, but a small minority of inherited forms allow gene identification which, together with neuropathology, yields important clues about the wider causes. Environmental and metabolic risk factors, including inflammation and vascular impairment, play a role in disease onset and progression. While neuronal atrophy and a loss of synapses occur throughout the cerebral cortex, we lack a full understanding of how this arises. The known hallmarks of Alzheimer's disease include amyloid- β plaques and neurofibrillary tau tangles and while extensive research has been carried out throughout the past few decades, the exact role of these protein aggregates in the disease remains elusive. In this chapter, we discuss mechanisms that have been implicated, including inflammation, mitochondrial dysfunction, oxidative stress and changes in protein clearance.

Keywords: amyloid- β plaques; etiology of Alzheimer's disease; dementia; neurodegeneration in Alzheimer's disease; neurofibrillary tau tangles

In: *Alzheimer's Disease: Drug Discovery*. Huang X (Editor). Exon Publications, Brisbane, Australia. ISBN: 978-0-6450017-0-9; Doi: <https://doi.org/10.36255/exonpublications.alzheimersdisease.2020>

Copyright: The Authors.

License: This open access article is licenced under Creative Commons Attribution-NonCommercial 4.0 International (CC BY-NC 4.0) <https://creativecommons.org/licenses/by-nc/4.0/>

INTRODUCTION

Around 50 million people worldwide suffer from dementia (1). About two thirds have Alzheimer's disease (AD) (2), an irreversible neurodegenerative disorder involving a decline in memory and executive function, and personality change (3). It is named after Alois Alzheimer who first characterized AD in 1906 (4). AD results in synapse loss and neuronal atrophy predominately throughout the hippocampus and cerebral cortex. It is characterized by amyloid plaques and neurofibrillary tau tangles (NFTs), aggregates of misfolded proteins, throughout the brain. Both genetics and environmental factors are believed to play a role in AD. While there are a small number of cases due to dominant genetic mutations (5–7), a majority of AD cases are sporadic and have no single genetic cause. Environmental and metabolic risk factors such as diabetes, cerebrovascular disease, poor diet, head injury and stress are linked to increased dementia risk. The leading hypothesis as to how AD begins and progresses, the amyloid hypothesis, though quite widely accepted, leaves many questions. In particular it remains unclear “what is the best drug target?” and “what lies upstream of the rise in amyloid- β ($A\beta$) in sporadic cases?” We still lack a fundamental understanding of how AD comes to fruition, and therapies to help individuals fight the disease. AD is a chronic disease manifesting as loss of memory, language, cognition and problem-solving skills, changes in behavior and ultimately death. While the primary signs are memory loss and executive dysfunction, they are often preceded by changes in language and vision (8). Additionally, not all types of memory are equally affected. People with AD have severely impaired episodic, semantic and working memory, yet long-term memory, such as procedural memory, tends to remain intact (9, 10). Clinically, AD is classified into seven stages (Table 1) (11). Patients often die 3–10 years after onset of symptoms (12) with complications arising from immobility, such as pneumonia or blood clots (13, 14).

TABLE 1
The seven clinical stages of Alzheimer's disease (Global Deterioration Scale) (11)

Symptoms and characteristics	
Stage 1	Persons appear cognitively normal, but pathological changes are happening in the brain.
Stage 2	Prodromal stage: mild memory loss, but generally this is indistinguishable from normal forgetfulness.
Stage 3	Progression into mild cognitive impairment (MCI). Individuals may get lost or have difficulty in finding correct wording.
Stage 4	Moderate dementia; poor short-term memory. Individuals forget some of their personal history.
Stage 5	Cognition continues to decline and at this point individuals need help in their daily lives. They suffer from confusion and forget many personal details.
Stage 6	Severe dementia. Requiring constant supervision and care. Patients fail to recognize many of their family and friends and have personality changes.
Stage 7	Individuals are nearing death. They show motor symptoms, have difficulty communicating, are incontinent and require assistance in feeding.

ETIOLOGY

Both genetic and environmental risk factors play a role in the manifestation of AD. The greatest risk factor is age. At age 65, the likelihood of having AD is about 3%, rising to over 30% by age 85 (15). The incidence of AD under the age of 65 is less certain, but estimates suggest that this age group accounts for around 3% of AD cases (15). Although overall numbers are increasing with the ageing population, age-specific incidence appears to be falling in several countries (16–18).

AD can be classified by when the disease manifests, and whether it is inherited. Early-onset Alzheimer's disease (EOAD) occurs before age 65, whereas late-onset Alzheimer's disease (LOAD) accounts for over 95% of cases (19) and manifests beyond age 65. Familial AD shows Mendelian (usually dominant) inheritance, while sporadic AD shows no simple familial link (20). Nearly all EOAD are familial as these cases are due to mutations in *APP*, *PSEN1* or *PSEN2*, and a vast majority of LOAD are sporadic. Genome wide association studies (GWAS) and sequencing have now provided more than 20 risk loci in total that contribute to sporadic cases (21), but often there is no identifiable genetic cause.

A β precursor protein

A β precursor protein (*APP*) was the first gene shown to have autosomal dominant mutations causing AD. As the precursor of the aggregated peptide in amyloid plaques, its discovery in 1991 by John Hardy and colleagues (5) led to the “amyloid hypothesis,” which states that the toxic build-up of A β starts a cascade of events, leading to neuronal death and disease (22, 23). There are now over 50 known *APP* mutations, accounting for approximately 10% of familial cases. Widely studied ones include the London (V717I) (24), Swedish (KM670/671NL) (25), Indiana (V717F) (26) and Arctic (E693G) (27) mutations, and most cluster around cleavage sites for β and γ -secretase (28). Research suggests that many of these mutations increase A β production, or the A β 42:40 ratio, leading to increased amyloid accumulation. In very rare instances, *APP* duplication or promoter mutations can cause AD (29, 30). Interestingly, studies have also found that there is an *APP* mutation (Icelandic—A673T) which lowers A β and protects against AD (31).

Presenilins

Presenilin 1 (*PSEN1*) and Presenilin 2 (*PSEN2*) encode the catalytic components of γ -secretase, an enzyme complex involved in *APP* processing (32). Presenilin mutations cause autosomal dominant AD, with *PSEN1* variants being the most commonly known Mendelian genetic cause, estimated to account for around 30–50% of familial EOAD cases (33, 34). Research shows that *PSEN1* and *PSEN2* mutations alter A β production, similar to *APP* mutations (35) but paradoxically tend to confer loss of function, raising questions as to how this fits the amyloid hypothesis (36, 37).

Other genetic risk factors

Other genes known to have variants associated with AD risk include *TREM2* (38), *APOE* (39), *CLU* (40–42), *SORL1* (43), *BIN1* (42) and *PICALM* (40, 42). *APOE*

(apolipoprotein E) is a protein involved in fat metabolism, and its E4 allele is the most common genetic risk factor for AD with an allele frequency of ~13.7% (44, 45). Heterozygosity for this allele increases the risk 3-fold (39). Although rarer, the variant *TREM2*^{R47H} (triggering receptor expressed on myeloid cells 2) has a similar effect size (46). *TREM2* is a receptor expressed on multiple cell types of the immune response, and its association supports a role for inflammation in AD pathogenesis.

Down syndrome

By age 65, up to 80% of Down syndrome (DS) individuals develop dementia (47). As with other instances of EOAD, amyloid and tau pathology begin much earlier than in LOAD, even at <40 years of age (48–50). DS results from the trisomy of chromosome 21, where the *APP* gene is located, and having three copies of this gene is sufficient to increase A β levels. However, the increased risk of developing the disease may also be due in part to triplication of other genes on chromosome 21 (47, 51, 52).

Inflammation

Sporadic AD often results from a combination of genetic and environmental risk factors, with cerebral hypoperfusion (53) and inflammation (54) being among the most common. Inflammation due to trauma, sepsis and infection has been linked to both short- and long-term cognitive impairment (55–57). Traumatic brain injury, and even bone fractures in the elderly, are implicated in dementia risk (58, 59). Higher levels of inflammatory markers such as interleukin 6 (IL-6) associate with greater risk of AD and vascular dementia (60). AD patients often have higher levels of certain inflammatory markers and activated microglia and astrocytes in the brain, which tend to surround plaques and tangles (61, 62). Finally, higher levels of these markers are associated with faster cognitive decline (63).

Cerebral, cardiovascular disease and diabetes

There is a strong link between vascular disease and dementia. Cardiovascular disease, including high blood pressure and heart attack, and cerebrovascular disease such as ischemia are associated with increased risk of AD (64). Metabolic and lifestyle risk factors for developing vascular diseases, including poor diet, obesity, high cholesterol and sedentary lifestyle, are also risk factors for dementia (65, 66). Poor diet and high cholesterol can produce metabolic changes both systemically and in the brain, and alter oxygen levels (67). Additionally, type 2 diabetes approximately doubles the risk for dementia (68–70).

Other environmental risk factors

The list of environmental and metabolic risk factors discussed here is not intended to be comprehensive, especially as the nature of epidemiology in populations with diverse genetics and lifestyle means that important mechanisms will not always

generate conclusive evidence. Other risk factors implicated include pollution, stress and heavy metal exposure (71–76). Many of these risk factors share some common characteristics with one another which can thus make it difficult to determine how their presence affects the brain. Some may act through similar mechanisms, such as inflammation or oxidative stress, which will be discussed later in this chapter.

NEUROPATHOLOGY

AD is characterised by synapse loss, followed by the atrophy of neurons throughout the cerebral cortex, with the medial temporal lobe being the most severely affected (77–79). Pathology appears to start within the hippocampus and entorhinal regions and spreads subsequently throughout the fronto-temporal cortices. It reaches as far as the striatum and thalamus, usually with sparing of the cerebellum (80–83). On a macroscale level, MRI scans show shrinkage of these regions (84). In particular, pyramidal cells of the CA1 of the hippocampus are vulnerable to morphological changes and cell death, consistent with the main symptom of memory loss (85, 86). The appearance of A β plaques and NFTs precedes clinical symptoms suggesting that by symptom onset, there have been years of pathological changes making early intervention difficult.

A β plaques

Senile plaques are primarily made of a variety of 36–43 residue-long amyloid peptides that undergo fibrilization to form A β sheets that are resistant to degradation (87). They often co-localize with neuronal debris and activated microglia and astrocytes (88), and first appear in the frontal, temporal and occipital lobes of the neocortex. They spread throughout neocortical areas as well as the hippocampal formation and entorhinal region, and eventually spread further throughout the cerebral cortex to the striatum and thalamus (83) (Figure 1). Amyloid pathology appears to precede that of tau, with NFTs only being found in regions where amyloid was already present. Numerous studies have shown that cognitively unimpaired elderly individuals can also have significant A β deposition (89–91), while on the contrary, others have reported a correlation of deposition to cognitive decline (92) and dementia severity (93). A recent study has more specifically shown that differences in A β oligomer concentration may be a better correlate of disease (94, 95). It is likely that differences in methodology are responsible for the varying conclusions from these studies. It has also been suggested that cognitively normal persons with high plaque levels may have “prodromal” disease, with A β pathology that precedes cognitive changes (96, 97).

Neuronal fibrillary tau tangles

NFTs are intraneuronal aggregates of hyperphosphorylated tau protein, encoded by the microtubule associated protein tau (*MAPT*) gene (98) (Figure 1). NFTs are

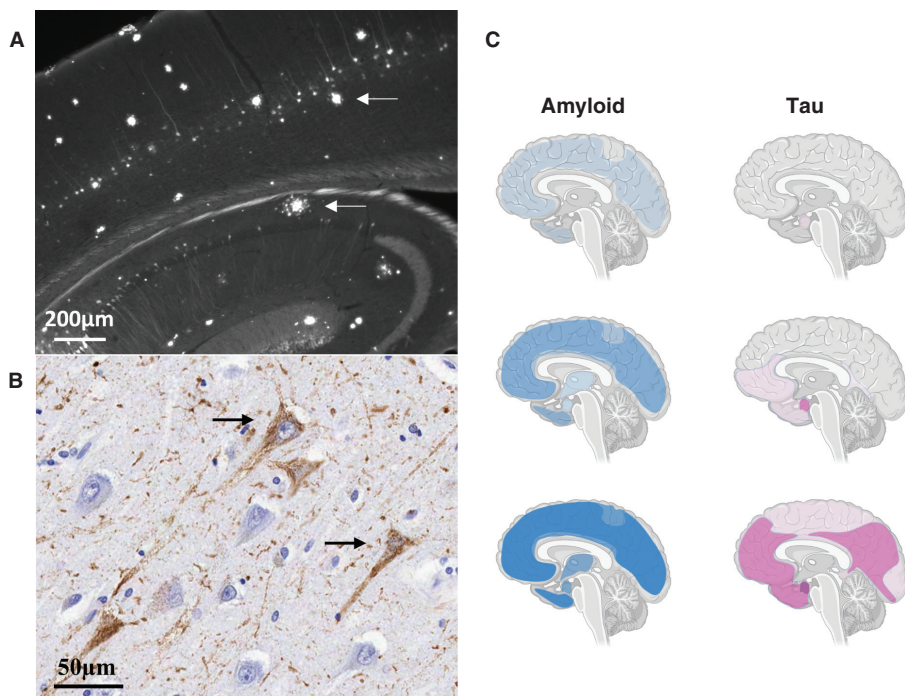


Figure 1. Amyloid and tau pathology. (A) Thioflavin S staining of A β plaques in the cortex of a CRND8 APP transgenic mouse. (B) AT8 staining of neurofibrillary tau tangles (NFTs) within an aged human CA1 region of the hippocampus. (C) The spread of amyloid and tau pathology throughout the brain during AD, adapted from Braak and Braak 1991 (83).

composed of paired helical fragments (PHFs) of tau fibrils approximately 20 nm in diameter (Figure 2). Like plaques, they spread throughout the brain as disease progresses, beginning near the entorhinal cortex. Braak staging is commonly used as a means of defining the progression of disease as determined by tau pathology. In stages I–II, tangles appear in the trans-entorhinal region; in stages III–IV, tangles have spread to the limbic system and start to show in the neocortex; in stages V–VI, pathology is present throughout the neocortex (83) (Figure 1). In addition to AD, several other neurodegenerative diseases are classified as tauopathies due to the presence of NFTs; these include Parkinson’s disease, progressive supranuclear palsy, corticobasal degeneration and frontotemporal dementia (FTD) (99). While aggregates of amyloid and tau have both been associated with neuronal loss and toxicity, they have a poor correlation with cognitive decline as AD progresses. On the contrary, the loss of synapses is one of the strongest correlates to cognitive decline in AD (100). Familial cases and PET imaging have allowed us to identify changes in both A β and tau prior to changes in brain structure and symptom onset (101). A combination of psychological and cognitive testing, scans and CSF and blood tests (to rule out other neurological disorders) are required to obtain the diagnosis of AD. Ultimately though, definitive confirmation of the disease requires post-mortem histopathology.

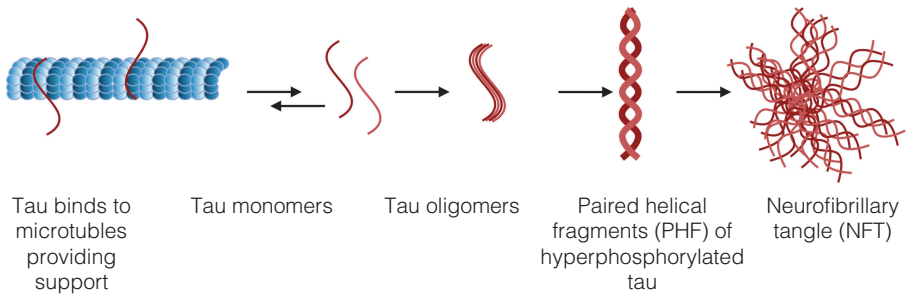


Figure 2. Microtubule-associated protein Tau (MAPT) aggregation results in the accumulation of neurofibrillary tau tangles (NFTs). Tau is believed to play a role in the stabilization of microtubules. Hyperphosphorylated tau polymerization leads to the creation of insoluble paired helical fragments (PHFs), which further aggregate into NFTs.

PATHOGENESIS

The mechanism of AD pathology and neuronal loss remains elusive. The roles of both A β and tau have been extensively researched in the past few decades, yet we are still unsure of their role in disease. A variety of mechanisms have been proposed to explain what occurs in the pathogenesis of AD. It is possible that different combinations of risk factors in different patients activate the disease in different ways, and that these converge on a common pathway of degeneration.

A β and APP

The amyloid hypothesis remains the dominant hypothesis in AD research due to the causal mutations found in both *APP* and presenilin genes. APP is processed via either the amyloidogenic or non-amyloidogenic pathway. For A β , APP is sequentially cleaved by the β - and γ -secretases, releasing the peptide into the cytosol (Figure 3). Functions of APP and A β are largely unknown, but they are thought to play a role in signal transduction for neuronal development, growth and survival (102, 103). While genetic mutations may explain A β accumulation in EOAD, it is still unclear how this occurs in LOAD. A β accumulation has been proposed to cause neuronal death via a number of mechanisms, including excitotoxicity, synaptic disruption, oxidative stress and mitochondrial dysfunction. Excitotoxicity can occur when NMDA receptors are continually activated, either by A β directly or by a downstream mechanism. In conjunction with synapse loss, both AD patients and animal models show reductions in the synaptic proteins synaptophysin and PSD-95 (104–108). A β oligomers accumulating in an AD brain (109) may be even more toxic than fibrils or plaques. Soluble oligomers appear to amass in a different manner compared to plaques and appear early in pathogenesis (110). Oligomers can disrupt cognitive function (111) and inhibit long-term potentiation (LTP) (112) *in vivo*, and can be neurotoxic (113) *in vitro*. Interestingly, oligomers tend to cluster near synapses (114) and can induce synapse loss and dysfunction (115). It has also been suggested that changes in another APP

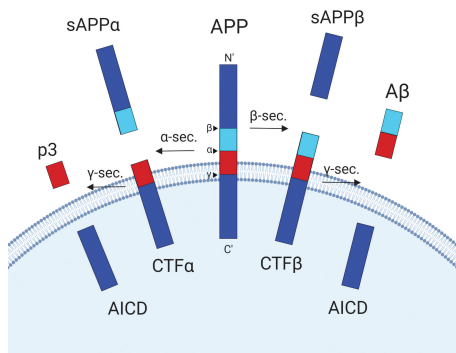


Figure 3. Post-translational processing of A β precursor protein (APP) is thought to occur at the cell surface or within endosomes. It includes cleavage by either α - then γ -secretase (non-amyloidogenic), or β - then γ -secretase (amyloidogenic pathway).

processing product could be a contributor to AD (103). Though many APP mouse models present with aspects of AD pathology, most fail to fully recapitulate the neurodegeneration seen in the human AD brain. While this most likely reflects inter-species differences, it also raises questions about the relative importance of APP/A β in driving dementia (116).

NFTs and Tau

While no *MAPT* mutations are associated with AD, causal mutations in tau have been found for other neurodegenerative diseases such as FTD, suggesting that tau dysfunction and aggregation can be neurotoxic. Tau's major role is thought to be that of a cytoskeletal protein, interacting with tubulin to help assemble and stabilize microtubules (117). In humans there are six isoforms of tau generated by alternative splicing of exons 2, 3 and 10. The incorporation of exon 10 leads to four microtubule-binding repeats (4R tau) instead of three (3R tau), altering how tightly the protein binds to microtubules and its propensity to aggregate (118). Healthy adult humans express similar amounts of 3R and 4R tau. Research has shown that the ratio between the two may impact disease, with higher 4R isoforms leading to greater degeneration. In AD, there is a higher ratio of 4R to 3R, and reported downstream consequences include transcriptional alterations in the Wnt signaling pathway (119) and altered axonal transport (120). Prior to NFT formation, tau becomes hyperphosphorylated, and tau phosphorylation not only plays a large role in regulating tau function, but could be the key change resulting in the accumulation, and potential toxicity, of this protein. In fact, multiple tauopathy mutations cause tau to be more readily phosphorylated (117).

Mutant tau mouse models have shown that mutations in this gene can result in severe neurological phenotypes (121, 122). Tau has been hypothesized to induce neurotoxicity via loss of function, gain of function and/or mis-localization. Loss of function of tau occurs when tau is no longer able to stabilize microtubules having an impact on neuronal cytoskeleton, and similarly could lead to deficiencies in axonal transport (123, 124). Higher levels of tau have also been shown to

inhibit vesicle and organelle trafficking, including those carrying APP, and increase levels of oxidative stress (125), as well as have an effect on axonal transport (126). The mis-localization of tau to dendritic spines has been shown to effect cognition and synapses *in vivo* (127, 128). As with APP, it remains unclear as to exactly how tau influences disease progression, but interestingly, A β induced toxicity and impairment in LTP has been found to be a requirement for the presence of endogenous tau (129, 130). It has also been suggested that tau and A β work together to result in transcriptional deficits (131) and synaptic changes (132) in AD.

Mitochondrial dysfunction and oxidative stress

One of the many processes that is compromised in AD is mitochondrial function. Alterations in mitochondrial morphology, number and transport, reduced cytochrome oxidase activity, deficiencies in metabolic proteins, changes in mitochondrial membrane potential and an increase in oxidative stress have been observed in AD (133, 134). Neurons are highly dependent on mitochondria, and mitochondria accumulate at synapses, helping to power their high metabolic demand. The high level of ROS production which occurs at synapses, in conjunction with insufficient antioxidants, can lead to oxidative stress (134). In addition, the brain is composed of high levels of cholesterol, which are also very vulnerable to oxidative damage (135). Thus, the high energy demands of the brain and its high lipid concentration naturally put it at risk for oxidative damage. Rather than aging driving amyloid pathology, as in the case of the amyloid hypothesis, the mitochondrial cascade hypothesis proposes that genetic and environmental factors determine the rate of mitochondrial decline, which in turn determines the rate of aging and subsequently AD (133). In terms of EOAD, APP or A β induces mitochondrial deficits, inducing an increase in the rate of aging, thus making some people susceptible to AD. This has been suggested as a potential link between EOAD and LOAD pathogenesis (136). Supporting this hypothesis, Thy-1-APP mice show reduced mitochondrial membrane potential and ATP synthesis and increased ROS production (137). Similarly, transgenic APP mice have shown an increase in A β within synaptic mitochondria, leading to dysfunction and oxidative stress prior to plaque accumulation (138). Paradoxically, oxidative stress, a by-product of mitochondrial deficiency, has been known to affect β -secretase activity (139), which in turn could alter A β production.

Insulin

Insulin resistance and a decrease in insulin receptors have been observed in the AD brain (140). Late stages of diabetes also result in insulin resistance in the brain. As cells are heavily dependent upon glucose metabolism for energy production, this can lead to energy deficiencies, potentially leading to oxidative stress. It has also been shown that insulin plays a role in neurotransmission (141) and can be neuroprotective during insults such as ischemia (142). Additionally, it has been reported that insulin and metabolic inhibitors result in increased levels of β -secretase in both wild-type and Tg2576 mice (an APP transgenic model). In Tg2576 mice, this also resulted in an increase in A β levels (143). Yet, as others report a protective role of insulin, it is likely that there is a certain level of this hormone which allows the brain to function optimally.

Hypoglycemia and vascular dysfunction

In addition to insulin resistance, the link between diabetes and AD could be due to changes in metabolic proteins, glucose receptors/transporters or even hypoglycemia due to over-medication. Glucose metabolism decreases in the normal aging brain (144) and even further in the AD brain (145). It has also been reported that there is a decline in the expression of glucose transporter at the blood brain barrier (BBB) in both AD patients and animal models of AD (146, 147), as well as in aged wild-type mice (147, 148). In addition, insulin-induced hypoglycemia has also been shown to cause neuronal death *in vitro* and *in vivo* (149). Glucose deprivation can elevate tau levels *in vitro* (150), and hypoglycemia has also been linked to increases in oxidative stress (151). Hypoglycemia could also be the link between cardiovascular and cerebral-vascular diseases and dementia, but whether it be hypoglycemia, hypoxia, a change in another blood component or a combination of these which increases one's risk of disease is still unknown. Finally, abnormal angiogenesis and alterations of vasculature, including changes in blood flow, have been shown in AD patients and animal models of the disease (152–154).

Inflammation

The role of inflammation is a more recent topic of interest in the AD field. As discussed previously, people with inflammation are more likely to develop dementia, and dementia patients with higher levels of inflammatory markers tend to deteriorate more rapidly. Studies in animal models have shown that inflammation can result in cognitive impairment (155), as well as neuronal damage and synaptic loss *in vivo* and *in vitro* (156–159). Although inflammation and the activation of microglia are thought to play a neuroprotective role in acute circumstances, in the long term, this may lead to neurotoxicity, and an increase in A β load (155, 160, 161). A β itself is thought to activate microglia, attracting them to plaques and enhancing phagocytosis (162–164). Potentially, microglial response to A β is protective, but after chronic activation, the microglia begin to play a detrimental role, resulting in a feed-forward loop of degradation (54). Similarly, it has been shown that increased ROS levels increase inflammatory markers, and that immune cells influence the production of ROS (165–168), demonstrating the complex interplay between A β , oxidative stress and inflammation.

Tau pathology also appears to be influenced by (169, 170), and have an effect upon (171, 172), inflammation. Research looking at the ability of microglia to phagocytose tau aggregates is conflicting, potentially due to microglia playing an initial role in clearance, but losing their ability to maintain this over extended periods (173). And finally, it has been reported that altering expression of TREM2, which plays a role in inflammation, may have an effect on A β levels and plaque-associated macrophages (174).

Ubiquitin-proteasome system

The ubiquitin-proteasome system (UPS) is involved in the degradation of misfolded and excess proteins. It is particularly important for synapse function, where there is high protein turnover (175). Proteins to be degraded go through an

enzymatic process where they are labelled with a polyubiquitin chain which is recognized by the proteasome (176), and subsequently broken down. The proteasome targets monomeric proteins, so is not thought to break down plaques or tangles, but both have been shown to potentially inhibit proteasome activity (177). This could lead to a toxic build-up of excess and misfolded proteins in the brain, and more specifically synapses.

Autophagy lysosome pathway

Autophagy and lysosomal dysfunction are also proposed mechanisms of AD pathogenesis. Autophagy is involved in tau clearance (178), and plays a role in both the generation and clearance of A β . APP amyloidogenic processing involves trafficking through the endo-lysosomal pathway (179). Several genes implicated in AD including *BIN1*, *SORL1* and *PICALM* are involved in endosomal recycling, and studies have reported that each may directly play a role in APP endosomal processing (95, 180, 181).

Cholinergic hypothesis

The cholinergic hypothesis was one of the first proposed theories on the manifestation of AD (182, 183). This came to fruition due to abnormal levels of acetylcholine in the AD brain. Cholinergic neurons of the basal forebrain are one of the earliest affected by AD and there is a decrease in choline acetyltransferase (ChAT) transcription and activity in remaining neurons. Studies have also shown a relationship between acetylcholinesterase (AChE) and A β accumulation (182). However, as the AD field has moved forward there has been difficulty in linking acetylcholine with other AD pathologies. Indeed, pyramidal neurons are lost in greatest numbers in regions with plaques and tangles and these are, for the most part, glutamatergic neurons (184).

CONCLUSION

Although we have amassed a vast amount of knowledge in the search for a central, unifying mechanism behind dementia and AD, we are still lacking suitable therapies to help slow down the progression of disease. The amyloid hypothesis remains the dominant theory, yet drugs aimed at lowering A β levels have been largely unsuccessful. The possibility of NFT and plaque-load being correlative rather than causative with disease progression is entirely possible. There is much overlap between many of the risk factors, both genetic and environmental, and the known pathogenesis, highlighting the complexity of dementia. Similarly, we lack a firm understanding of how familial EOAD and sporadic LOAD ultimately produce the same neurodegenerative outcome. By enhancing our understanding of AD etiology, pathology and pathogenesis, we hope to one day find an effective therapy.

Acknowledgements: Olivia Sheppard is currently funded by an Alzheimer's Research UK project grant. The authors thank Dr. Robert Adalbert for contributing the A β plaque image in Figure 1A. The authors also thank Dr. Antonina Kouli,

Dr. Caroline Williams-Gray and the Cambridge Brain Bank for the NFT image in Figure 1B. The Cambridge Brain Bank is supported by the NIHR Cambridge Biomedical Research Centre.

Conflict of Interest: The authors declare no potential conflicts of interest with respect to research, authorship and/or publication of this chapter.

Copyright and Permission: To the best of our knowledge, the materials included in this chapter do not violate copyright laws. All original sources have been appropriately acknowledged and/or referenced. Where relevant, appropriate permissions have been obtained from the original copyright holder(s).

REFERENCES

1. World Health Organization. Risk reduction of cognitive decline and dementia: WHO guidelines. Geneva: World Health Organization; 2019.
2. World Health Organization. Global action plan on the public health response to dementia 2017–2025. Geneva: World Health Organization; 2017.
3. Wisniewski T, editor. Alzheimer's disease. Brisbane: Codon Publications; 2019. <https://doi.org/10.15586/alzheimersdisease.2019>
4. Hippus H, Neundörfer G. The discovery of Alzheimer's disease. *Dialogues Clin Neurosci*. 2003;5(1):101–8.
5. Goate A, Chartier-Harlin MC, Mullan M, Brown J, Crawford F, Fidani L, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature*. 1991 Feb 21;349(6311):704–6. <http://dx.doi.org/10.1038/349704a0>
6. Sherrington R, Rogaev EI, Liang Y, Rogaeva EA, Levesque G, Ikeda M, et al. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature*. 1995;375(6534):754–60. <http://dx.doi.org/10.1038/37554a0>
7. Levy-Lahad E, Wasco W, Poorkaj P, Romano DM, Oshima J, Pettingell WH, et al. Candidate gene for the chromosome 1 familial Alzheimer's disease locus. *Science* (80-). 1995;269(5226):973–7. <http://dx.doi.org/10.1126/science.7638622>
8. Schachter AS, Davis KL. Alzheimer's disease. *Dialogues Clin Neurosci*. 2000 Jun;2(2):91–100.
9. Gold CA, Budson AE. Memory loss in Alzheimer's disease: Implications for development of therapeutics. *Expert Rev Neurother*. 2008 Dec 9;8(12):1879–91. <http://dx.doi.org/10.1586/14737175.8.12.1879>
10. Budson AE, Price BH. Memory dysfunction. *N Engl J Med*. 2005 Feb 17;352(7):692–9. <http://dx.doi.org/10.1056/NEJMra041071>
11. Reisberg B, Ferris S, De Leon M, Crook T. The Global Deterioration Scale for assessment of primary degenerative dementia. *Am J Psychiatry*. 1982 Sep;139(9):1136–9. <http://dx.doi.org/10.1176/ajp.139.9.1136>
12. Zanetti O, Solerte SB, Cantoni F. Life expectancy in Alzheimer's disease (AD). *Arch Gerontol Geriatr*. 2009 Jan;49:237–43. <http://dx.doi.org/10.1016/j.archger.2009.09.035>
13. Alzheimer's Association. 2016 Alzheimer's disease facts and figures. *Alzheimer's Dement*. 2016 Apr;12(4):459–509. <http://dx.doi.org/10.1016/j.jalz.2016.03.001>
14. Burns A, Jacoby R, Luthert P, Levy R. Cause of death in Alzheimer's disease. *Age Ageing*. 1990 Sep;19(5):341–4.
15. Alzheimer's Association. 2019 Alzheimer's disease facts and figures. *Alzheimer's Dement*. 2019 Mar;15(3):321–87. <http://dx.doi.org/10.1016/j.jalz.2019.01.010>
16. Matthews FE, Arthur A, Barnes LE, Bond J, Jagger C, Robinson L, et al. A two-decade comparison of prevalence of dementia in individuals aged 65 years and older from three geographical areas of England: Results of the cognitive function and ageing study I and II. *Lancet*. 2013;382(9902):1405–12. [http://dx.doi.org/10.1016/S0140-6736\(13\)61570-6](http://dx.doi.org/10.1016/S0140-6736(13)61570-6)

17. Matthews FE, Stephan BCM, Robinson L, Jagger C, Barnes LE, Arthur A, et al. A two decade dementia incidence comparison from the cognitive function and ageing studies I and II. *Nat Commun*. 2016;7:11398. <http://dx.doi.org/10.1038/ncomms11398>
18. Schrijvers EMC, Verhaaren BFJ, Koudstaal PJ, Hofman A, Ikram MA, Breteler MMB. Is dementia incidence declining? Trends in dementia incidence since 1990 in the Rotterdam study. *Neurology*. 2012;78(19):1456–63. <http://dx.doi.org/10.1212/WNL.0b013e3182553be6>
19. Masters CL, Bateman R, Blennow K, Rowe CC, Sperling RA, Cummings JL. Alzheimer's disease. *Nat Rev Dis Prim*. 2015 Dec 15;1(1):15056. <http://dx.doi.org/10.1038/nrdp.2015.56>
20. Talwar P, Sinha J, Grover S, Rawat C, Kushwaha S, Agarwal R, et al. Dissecting complex and multifactorial nature of Alzheimer's disease pathogenesis: A clinical, genomic, and systems biology perspective. *Mol Neurobiol*. 2016;53(7):4833–64. <http://dx.doi.org/10.1007/s12035-015-9390-0>
21. Cuyvers E, Sleegers K. Genetic variations underlying Alzheimer's disease: Evidence from genome-wide association studies and beyond. *Lancet Neurol*. 2016;15(8):857–68. [http://dx.doi.org/10.1016/S1474-4422\(16\)00127-7](http://dx.doi.org/10.1016/S1474-4422(16)00127-7)
22. Hardy J, Higgins G. Alzheimer's disease: The amyloid cascade hypothesis. *Science* (80-). 1992 Apr 10;256(5054):184–5. <http://dx.doi.org/10.1126/science.1566067>
23. Hardy J, Allsop D. Amyloid deposition as the central event in the aetiology of Alzheimer's disease. *Trends Pharmacol Sci*. 1991 Jan;12:383–8. [http://dx.doi.org/10.1016/0165-6147\(91\)90609-V](http://dx.doi.org/10.1016/0165-6147(91)90609-V)
24. Eckman CB, Mehta ND, Crook R, Perez-tur J, Prihar G, Pfeiffer E, et al. A new pathogenic mutation in the APP gene (I716V) increases the relative proportion of A beta 42(43). *Hum Mol Genet*. 1997 Nov;6(12):2087–9. <http://dx.doi.org/10.1093/hmg/6.12.2087>
25. Mullan M, Crawford F, Axelman K, Houlden H, Lilius L, Winblad B, et al. A pathogenic mutation for probable Alzheimer's disease in the APP gene at the N-terminus of beta-amyloid. *Nat Genet*. 1992 Aug;1(5):345–7. <http://dx.doi.org/10.1038/ng0892-345>
26. Murrell J, Farlow M, Ghetti B, Benson M. A mutation in the amyloid precursor protein associated with hereditary Alzheimer's disease. *Science* (80-). 1991 Oct 4;254(5028):97–9. <http://dx.doi.org/10.1126/science.1925564>
27. Nilsberth C, Westlind-Danielsson A, Eckman CB, Condron MM, Axelman K, Forsell C, et al. The "Arctic" APP mutation (E693G) causes Alzheimer's disease by enhanced A beta protofibril formation. *Nat Neurosci*. 2001 Sep 20;4(9):887–93. <http://dx.doi.org/10.1038/nn0901-887>
28. De Jonghe C, Esselens C, Kumar-Singh S, Craessaerts K, Serneels S, Checler F, et al. Pathogenic APP mutations near the gamma-secretase cleavage site differentially affect Abeta secretion and APP C-terminal fragment stability. *Hum Mol Genet*. 2001 Aug 1;10(16):1665–71. <http://dx.doi.org/10.1093/hmg/10.16.1665>
29. McNaughton D, Knight W, Guerreiro R, Ryan N, Lowe J, Poulter M, et al. Duplication of amyloid precursor protein (APP), but not prion protein (PRNP) gene is a significant cause of early onset dementia in a large UK series. *Neurobiol Aging*. 2012 Feb;33(2):426.e13–21. <http://dx.doi.org/10.1016/j.neurobiolaging.2010.10.010>
30. Theuns J, Brouwers N, Engelborghs S, Sleegers K, Bogaerts V, Corsmit E, et al. Promoter mutations that increase amyloid precursor-protein expression are associated with Alzheimer disease. *Am J Hum Genet*. 2006 Jun;78(6):936–46. <http://dx.doi.org/10.1086/504044>
31. Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, et al. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. *Nature*. 2012 Aug 2;488(7409):96–9. <http://dx.doi.org/10.1038/nature11283>
32. Brunkan AL, Goate AM. Presenilin function and gamma-secretase activity. *J Neurochem*. 2005 May;93(4):769–92. <http://dx.doi.org/10.1111/j.1471-4159.2005.03099.x>
33. Singleton AB, Hall R, Ballard CG, Perry RH, Xuereb JH, Rubinsztein DC, et al. Pathology of early-onset Alzheimer's disease cases bearing the Thr113-114ins presenilin-1 mutation. *Brain*. 2000 Dec 1;123 Pt 12(12):2467–74. <http://dx.doi.org/10.1093/brain/123.12.2467>
34. Cruts M, van Duijn CM, Backhovens H, Van den Broeck M, Wehnert A, Serneels S, et al. Estimation of the genetic contribution of presenilin-1 and -2 mutations in a population-based study of presenile Alzheimer disease. *Hum Mol Genet*. 1998 Jan;7(1):43–51. <http://dx.doi.org/10.1093/hmg/7.1.43>
35. Borchelt DR, Thinakaran G, Eckman CB, Lee MK, Davenport F, Ratovitsky T, et al. Familial Alzheimer's disease-linked presenilin 1 variants elevate A beta 1–42/1–40 ratio in vitro and in vivo. *Neuron*. 1996 Nov;17(5):1005–13. [http://dx.doi.org/10.1016/S0896-6273\(00\)80230-5](http://dx.doi.org/10.1016/S0896-6273(00)80230-5)

36. De Strooper B. Loss-of-function presenilin mutations in Alzheimer disease. *EMBO Rep.* 2007 Feb;8(2):141–6. <http://dx.doi.org/10.1038/sj.embor.7400897>
37. Xia D, Watanabe H, Wu B, Lee SH, Li Y, Tsvetkov E, et al. Presenilin-1 knockin mice reveal loss-of-function mechanism for familial Alzheimer's disease. *Neuron.* 2015 Mar 4;85(5):967–81. <http://dx.doi.org/10.1016/j.neuron.2015.02.010>
38. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, et al. TREM2 variants in Alzheimer's disease. *N Engl J Med.* 2013 Jan 10;368(2):117–27. <http://dx.doi.org/10.1056/NEJMoa1211851>
39. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science (80-).* 1993;261(5123):921–3. <http://dx.doi.org/10.1126/science.8346443>
40. Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, et al. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet.* 2009 Oct 6;41(10):1088–93.
41. Lambert J-C, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, et al. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet.* 2009 Oct 6;41(10):1094–9. <http://dx.doi.org/10.1038/ng.439>
42. Lee JH, Cheng R, Barral S, Reitz C, Medrano M, Lantigua R, et al. Identification of novel loci for Alzheimer disease and replication of CLU, PICALM, and BIN1 in Caribbean Hispanic individuals. *Arch Neurol.* 2011 Mar 1;68(3):320–8. <http://dx.doi.org/10.1001/archneurol.2010.292>
43. Rogaeva E, Meng Y, Lee JH, Gu Y, Kawarai T, Zou F, et al. The neuronal sortilin-related receptor SORL1 is genetically associated with Alzheimer disease. *Nat Genet.* 2007 Feb 14;39(2):168–77.
44. Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, et al. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. *JAMA.* 1997 Oct 22;278(16):1349–56. <http://dx.doi.org/10.1001/jama.278.16.1349>
45. Liu C-C, Kanekiyo T, Xu H, Bu G. Apolipoprotein E and Alzheimer disease: Risk, mechanisms and therapy. *Nat Rev Neurol.* 2013 Feb 8;9(2):106–18. <http://dx.doi.org/10.1038/nrneurol.2012.263>
46. Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, et al. Variant of TREM2 associated with the risk of Alzheimer's disease. *N Engl J Med.* 2013 Jan 10;368(2):107–16. <http://dx.doi.org/10.1056/NEJMoa1211103>
47. Wiseman FK, Al-Janabi T, Hardy J, Karmiloff-Smith A, Nizetic D, Tybulewicz VLJ, et al. A genetic cause of Alzheimer disease: Mechanistic insights from Down syndrome. *Nat Rev Neurosci.* 2015 Sep;16(9):564–74. <http://dx.doi.org/10.1038/nrn3983>
48. Mann DMA. Alzheimer's disease and Down's syndrome. *Histopathology.* 1988 Aug;13(2):125–37. <http://dx.doi.org/10.1111/j.1365-2559.1988.tb02018.x>
49. Allsop D, Haga SI, Haga C, Ikeda SI, Mann DMA, Ishii T. Early senile plaques in down's syndrome brains show a close relationship with cell bodies of neurons. *Neuropathol Appl Neurobiol.* 1989;15(6):531–42. <http://dx.doi.org/10.1111/j.1365-2990.1989.tb01252.x>
50. Abrahamson EE, Head E, Lott IT, Handen BL, Mufson EJ, Christian BT, et al. Neuropathological correlates of amyloid PET imaging in down syndrome. *Dev Neurobiol.* 2019;79(7):750–66. <http://dx.doi.org/10.1002/dneu.22713>
51. Kimura R, Kamino K, Yamamoto M, Nuripa A, Kida T, Kazui H, et al. The DYRK1A gene, encoded in chromosome 21 down syndrome critical region, bridges between β -amyloid production and tau phosphorylation in Alzheimer disease. *Hum Mol Genet.* 2007;16(1):15–23. <http://dx.doi.org/10.1093/hmg/ddl437>
52. Sheppard O, Plattner F, Rubin A, Slender A, Linehan JM, Brandner S, et al. Altered regulation of tau phosphorylation in a mouse model of down syndrome aging. *Neurobiol Aging.* 2012 Apr;33(4):828.e31–828.e44. <http://dx.doi.org/10.1016/j.neurobiolaging.2011.06.025>
53. Duncombe J, Kitamura A, Hase Y, Ihara M, Kalara RN, Horsburgh K. Chronic cerebral hypoperfusion: A key mechanism leading to vascular cognitive impairment and dementia. Closing the translational gap between rodent models and human vascular cognitive impairment and dementia. *Clin Sci.* 2017;131(19):2451–68. <http://dx.doi.org/10.1042/CS20160727>

54. Kinney JW, Bemiller SM, Murtishaw AS, Leisgang AM, Salazar AM, Lamb BT. Inflammation as a central mechanism in Alzheimer's disease. *Alzheimer's Dement Transl Res Clin Interv*. 2018 Jan 5;4(1):575–90. <http://dx.doi.org/10.1016/j.trci.2018.06.014>
55. Iwashyna TJ, Ely EW, Smith DM, Langa KM. Long-term cognitive impairment and functional disability among survivors of severe sepsis. *JAMA*. 2010 Oct 27;304(16):1787–94. <http://dx.doi.org/10.1001/jama.2010.1553>
56. Annane D, Sharshar T. Cognitive decline after sepsis. *Lancet Respir Med*. 2015;3(1):61–9. [http://dx.doi.org/10.1016/S2213-2600\(14\)70246-2](http://dx.doi.org/10.1016/S2213-2600(14)70246-2)
57. Benros ME, Sørensen HJ, Nielsen PR, Nordentoft M, Mortensen PB, Petersen L. The association between infections and general cognitive ability in young men—A nationwide study. *PLoS One*. 2015;10(5):1–13. <http://dx.doi.org/10.1371/journal.pone.0124005>
58. Fann JR, Ribe AR, Pedersen HS, Fenger-Grøn M, Christensen J, Benros ME, et al. Long-term risk of dementia among people with traumatic brain injury in Denmark: A population-based observational cohort study. *Lancet Psychiatry*. 2018;5(5):424–31. [http://dx.doi.org/10.1016/S2215-0366\(18\)30065-8](http://dx.doi.org/10.1016/S2215-0366(18)30065-8)
59. Tsai C-H, Chuang C-S, Hung C-H, Lin C-L, Sung F-C, Tang C-H, et al. Fracture as an independent risk factor of dementia: A nationwide population-based cohort study. *Medicine (Baltimore)*. 2014 Nov;93(26):e188. <http://dx.doi.org/10.1097/MD.0000000000000188>
60. Engelhart MJ, Geerlings MI, Meijer J, Kiliaan A, Ruitenberg A, van Swieten JC, et al. Inflammatory proteins in plasma and the risk of dementia. *Arch Neurol*. 2004 May 1;61(5):668. <http://dx.doi.org/10.1001/archneur.61.5.668>
61. Beach TG, Walker R, McGeer EG. Patterns of gliosis in alzheimer's disease and aging cerebrum. *Glia*. 1989;2(6):420–36. <http://dx.doi.org/10.1002/glia.440020605>
62. Navarro V, Sanchez-Mejias E, Jimenez S, Muñoz-Castro C, Sanchez-Varo R, Davila JC, et al. Microglia in Alzheimer's disease: Activated, dysfunctional or degenerative. *Front Aging Neurosci*. 2018 May 11;10(5):1005–13. <http://dx.doi.org/10.3389/fnagi.2018.00140>
63. Passamonti L, Tsvetanov KA, Jones PS, Bevan-Jones WR, Arnold R, Borchert RJ, et al. Neuroinflammation and functional connectivity in Alzheimer's disease: Interactive influences on cognitive performance. *J Neurosci*. 2019;39(36):7218–26. <http://dx.doi.org/10.1523/JNEUROSCI.2574-18.2019>
64. Stampfer MJ. Cardiovascular disease and Alzheimer's disease: Common links. *J Intern Med*. 2006 Sep;260(3):211–23. <http://dx.doi.org/10.1111/j.1365-2796.2006.01687.x>
65. Akbaraly TN, Singh-Manoux A, Dugravot A, Brunner EJ, Kivimäki M, Sabia S. Association of midlife diet with subsequent risk for dementia. *JAMA—J Am Med Assoc*. 2019;321(10):957–68. <http://dx.doi.org/10.1001/jama.2019.1432>
66. de Bruijn RFAG, Ikram MA. Cardiovascular risk factors and future risk of Alzheimer's disease. *BMC Med*. 2014 Nov 11;12(1):130. <http://dx.doi.org/10.1186/s12916-014-0130-5>
67. Uraga RM, Keller JN. Diet and age interactions with regards to cholesterol regulation and brain pathogenesis. *Curr Gerontol Geriatr Res*. 2010;2010(Vldl):219683. <http://dx.doi.org/10.1155/2010/219683>
68. Ott A, Stolk RP, van Harskamp F, Pols HAP, Hofman A, Breteler MMB. Diabetes mellitus and the risk of dementia: The Rotterdam study. *Neurology*. 1999 Dec 1;53(9):1937. <http://dx.doi.org/10.1212/WNL.53.9.1937>
69. Biessels GJ, Staekenborg S, Brunner E, Brayne C, Scheltens P. Risk of dementia in diabetes mellitus: A systematic review. *Lancet Neurol*. 2006 Jan;5(1):64–74. [http://dx.doi.org/10.1016/S1474-4422\(05\)70284-2](http://dx.doi.org/10.1016/S1474-4422(05)70284-2)
70. Butterfield DA, Di Domenico F, Barone E. Elevated risk of type 2 diabetes for development of Alzheimer disease: A key role for oxidative stress in brain. *Biochim Biophys Acta*. 2014 Sep;1842(9):1693–706. <http://dx.doi.org/10.1016/j.bbadis.2014.06.010>
71. Armstrong RA. Risk factors for Alzheimer's disease. *Folia Neuropathol*. 2019;57(2):87–105. <http://dx.doi.org/10.5114/fn.2019.85929>
72. Chen H, Kwong JC, Copes R, Tu K, Villeneuve PJ, van Donkelaar A, et al. Living near major roads and the incidence of dementia, Parkinson's disease, and multiple sclerosis: A population-based cohort study. *Lancet (London, England)*. 2017;389(10070):718–26. [http://dx.doi.org/10.1016/S0140-6736\(16\)32399-6](http://dx.doi.org/10.1016/S0140-6736(16)32399-6)

73. Justice NJ. The relationship between stress and Alzheimer's disease. *Neurobiol Stress*. 2018 Feb;8(April):127–33. <http://dx.doi.org/10.1016/j.ynstr.2018.04.002>
74. Neri LC, Hewitt D. Aluminium, Alzheimer's disease, and drinking water. *Lancet (London, England)*. 1991 Aug 10;338(8763):390. [http://dx.doi.org/10.1016/0140-6736\(91\)90531-5](http://dx.doi.org/10.1016/0140-6736(91)90531-5)
75. Forbes WF, McAiney CA. Aluminum and dementia. *Lancet (London, England)*. 1992 Sep 12; 340(8820):668–9. [http://dx.doi.org/10.1016/0140-6736\(92\)92198-O](http://dx.doi.org/10.1016/0140-6736(92)92198-O)
76. Cacciottolo M, Wang X, Driscoll I, Woodward N, Saffari A, Reyes J, et al. Particulate air pollutants, APOE alleles and their contributions to cognitive impairment in older women and to amyloidogenesis in experimental models. *Transl Psychiatry*. 2017;7(1):e1022. <http://dx.doi.org/10.1038/tp.2016.280>
77. Sheng M, Sabatini BL, Südhof TC. Synapses and Alzheimer's disease. *Cold Spring Harb Perspect Biol*. 2012 May 1;4(5):1–18. <http://dx.doi.org/10.1101/cshperspect.a005777>
78. Kashyap G, Bapat D, Das D, Gowaikar R, Amritkar RE, Rangarajan G, et al. Synapse loss and progress of Alzheimer's disease—A network model. *Sci Rep*. 2019;9(1):1–9. <http://dx.doi.org/10.1038/s41598-019-43076-y>
79. Chen M-K, Mecca AF, Naganawa M, Finnema SJ, Toyonaga T, Lin S-F, et al. Assessing synaptic density in Alzheimer disease with synaptic vesicle glycoprotein 2A positron emission tomographic imaging. *JAMA Neurol*. 2018;75(10):1215–24. <http://dx.doi.org/10.1001/jamaneurol.2018.1836>
80. Duara R, Loewenstein DA, Potter E, Appel J, Greig MT, Urs R, et al. Medial temporal lobe atrophy on MRI scans and the diagnosis of Alzheimer disease. *Neurology*. 2008 Dec 9;71(24):1986–92. <http://dx.doi.org/10.1212/01.wnl.0000336925.79704.9f>
81. McDonald CR, McEvoy LK, Gharapetian L, Fennema-Notestine C, Hagler DJ, Holland D, et al. Regional rates of neocortical atrophy from normal aging to early Alzheimer disease. *Neurology*. 2009 Aug 11;73(6):457–65. <http://dx.doi.org/10.1212/WNL.0b013e3181b16431>
82. Mann DMA. The topographic distribution of brain atrophy in Alzheimer's disease. *Acta Neuropathol*. 1991 Dec;83(1):81–6. <http://dx.doi.org/10.1007/BF00294434>
83. Braak H, Braak E. Neuropathological staging of Alzheimer-related changes. *Acta Neuropathol*. 1991 Sep;82(4):239–59. <http://dx.doi.org/10.1007/BF00308809>
84. Perl DP. Neuropathology of Alzheimer's disease. *Mt Sinai J Med A J Transl Pers Med*. 2010 Jan;77(1): 32–42. <http://dx.doi.org/10.1002/msj.20157>
85. Braak E, Braak H. Alzheimer's disease: Transiently developing dendritic changes in pyramidal cells of sector CA1 of the Ammon's horn. *Acta Neuropathol*. 1997;93(4):323–5. <http://dx.doi.org/10.1007/s004010050622>
86. Masurkar AV. Towards a circuit-level understanding of hippocampal CA1 dysfunction in Alzheimer's disease across anatomical axes. *J Alzheimer's Dis Park*. 2018;8(1):100–6. <http://dx.doi.org/10.4172/2161-0460.1000412>
87. Thal DR, Walter J, Saido TC, Fändrich M. Neuropathology and biochemistry of A β and its aggregates in Alzheimer's disease. *Acta Neuropathol*. 2015 Feb 23;129(2):167–82. <http://dx.doi.org/10.1007/s00401-014-1375-y>
88. Ries M, Sastre M. Mechanisms of A β clearance and degradation by glial cells. *Front Aging Neurosci*. 2016;8(Jun):1–9. <http://dx.doi.org/10.3389/fnagi.2016.00160>
89. Aizenstein HJ, Nebes RD, Saxton JA, Price JC, Mathis CA, Tsopelas ND, et al. Frequent amyloid deposition without significant cognitive impairment among the elderly. *Arch Neurol*. 2008 Nov 10;65(11):1509. <http://dx.doi.org/10.1001/archneur.65.11.1509>
90. Dickson DW, Crystal HA, Mattiace LA, Masur DM, Blau AD, Davies P, et al. Identification of normal and pathological aging in prospectively studied nondemented elderly humans. *Neurobiol Aging*. 1992;13(1):179–89. [http://dx.doi.org/10.1016/0197-4580\(92\)90027-U](http://dx.doi.org/10.1016/0197-4580(92)90027-U)
91. Katzman R, Terry R, DeTeresa R, Brown T, Davies P, Fuld P, et al. Clinical, pathological, and neurochemical changes in dementia: A subgroup with preserved mental status and numerous neocortical plaques. *Ann Neurol*. 1988;23(2):138–44. <http://dx.doi.org/10.1002/ana.410230206>
92. Cummings B, Cotman C. Image analysis of β -amyloid load in Alzheimer's disease and relation to dementia severity. *Lancet*. 1995 Dec;346(8989):1524–8. [http://dx.doi.org/10.1016/S0140-6736\(95\)92053-6](http://dx.doi.org/10.1016/S0140-6736(95)92053-6)
93. Nielson KA, Cummings BJ, Cotman CW. Constructional apraxia in Alzheimer's disease correlates with neuritic neuropathology in occipital cortex. *Brain Res*. 1996;741(1–2):284–93. [http://dx.doi.org/10.1016/S0006-8993\(96\)00983-3](http://dx.doi.org/10.1016/S0006-8993(96)00983-3)

94. Esparza TJ, Zhao H, Cirrito JR, Cairns NJ, Bateman RJ, Holtzman DM, et al. Amyloid-beta oligomerization in Alzheimer dementia versus high-pathology controls. *Ann Neurol*. 2013 Jan;73(1):104–19. <http://dx.doi.org/10.1002/ana.23748>
95. Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med*. 2016 Nov;8(6):595–608. <http://dx.doi.org/10.15252/emmm.201606210>
96. Price JL, Morris JC. Tangles and plaques in nondemented aging and “preclinical” alzheimer's disease. *Ann Neurol*. 1999;45(3):358–68. [http://dx.doi.org/10.1002/1531-8249\(199903\)45:3%3C358::AID-ANA12%3E3.0.CO;2-X](http://dx.doi.org/10.1002/1531-8249(199903)45:3%3C358::AID-ANA12%3E3.0.CO;2-X)
97. Hulette CM, Welsh-Bohmer KA, Murray MG, Saunders AM, Mash DC, Mcintyre LM. Neuropathological and neuropsychological changes in “normal” aging. *J Neuropathol Exp Neurol*. 1998 Dec;57(12):1168–74. <http://dx.doi.org/10.1097/00005072-199812000-00009>
98. Binder LI, Guillozet-Bongaarts AL, Garcia-Sierra F, Berry RW. Tau, tangles, and Alzheimer's disease. *Biochim Biophys Acta—Mol Basis Dis*. 2005 Jan;1739(2–3):216–23. <http://dx.doi.org/10.1016/j.bbadis.2004.08.014>
99. Irwin DJ. Tauopathies as clinicopathological entities. *Parkinsonism Relat Disord*. 2016 Jan;22 Suppl 1(3):S29–33. <http://dx.doi.org/10.1016/j.parkreldis.2015.09.020>
100. Terry RD, Masliah E, Salmon DP, Butters N, DeTeresa R, Hill R, et al. Physical basis of cognitive alterations in Alzheimer's disease: Synapse loss is the major correlate of cognitive impairment. *Ann Neurol*. 1991 Oct;30(4):572–80. <http://dx.doi.org/10.1002/ana.410300410>
101. Thal DR, Attems J, Ewers M. Spreading of amyloid, tau, and microvascular pathology in alzheimer's disease: Findings from neuropathological and neuroimaging studies. *J Alzheimer's Dis*. 2014;42: S421–9. <http://dx.doi.org/10.3233/JAD-141461>
102. Dawkins E, Small DH. Insights into the physiological function of the β -amyloid precursor protein: Beyond Alzheimer's disease. *J Neurochem*. 2014 Jun;129(5):756–69. <http://dx.doi.org/10.1111/jnc.12675>
103. Zhang Y, Thompson R, Zhang H, Xu H. APP processing in Alzheimer's disease. *Mol Brain*. 2011 Nov;4(1):3. <http://dx.doi.org/10.1186/1756-6606-4-3>
104. Sza C-I, Troncoso JC, Kawas C, Mouton P, Price DL, Martin LJ. Loss of the presynaptic vesicle protein synaptophysin in hippocampus correlates with cognitive decline in Alzheimer disease. *J Neuropathol Exp Neurol*. 1997 Aug;56(8):933–44. <http://dx.doi.org/10.1097/00005072-199708000-00011>
105. Gyls KH, Fein JA, Yang F, Wiley DJ, Miller CA, Cole GM. Synaptic changes in Alzheimer's disease. *Am J Pathol*. 2004 Nov;165(5):1809–17. [http://dx.doi.org/10.1016/S0002-9440\(10\)63436-0](http://dx.doi.org/10.1016/S0002-9440(10)63436-0)
106. Mucke L, Masliah E, Yu GQ, Mallory M, Rockenstein EM, Tatsuno G, et al. High-level neuronal expression of $A\beta(1-42)$ in wild-type human amyloid protein precursor transgenic mice: Synaptotoxicity without plaque formation. *J Neurosci*. 2000;20(11):4050–8. <http://dx.doi.org/10.1523/JNEUROSCI.20-11-04050.2000>
107. Shao CY, Mirra SS, Sait HBR, Sacktor TC, Sigurdsson EM. Postsynaptic degeneration as revealed by PSD-95 reduction occurs after advanced $A\beta$ and tau pathology in transgenic mouse models of Alzheimer's disease. *Acta Neuropathol*. 2011;122(3):285–92. <http://dx.doi.org/10.1007/s00401-011-0843-x>
108. Harwell CS, Coleman MP. Synaptophysin depletion and intraneuronal $A\beta$ in organotypic hippocampal slice cultures from huAPP transgenic mice. *Mol Neurodegener*. 2016;11(1):44. <http://dx.doi.org/10.1186/s13024-016-0110-7>
109. Gong Y, Chang L, Viola KL, Lacor PN, Lambert MP, Finch CE, et al. Alzheimer's disease-affected brain: Presence of oligomeric A ligands (ADDLs) suggests a molecular basis for reversible memory loss. *Proc Natl Acad Sci*. 2003 Sep 2;100(18):10417–22. <http://dx.doi.org/10.1073/pnas.1834302100>
110. Viola KL, Klein WL. Amyloid β oligomers in Alzheimer's disease pathogenesis, treatment, and diagnosis. *Acta Neuropathol*. 2015 Feb;129(2):183–206. <http://dx.doi.org/10.1007/s00401-015-1386-3>
111. Cleary JP, Walsh DM, Hofmeister JJ, Shankar GM, Kuskowski MA, Selkoe DJ, et al. Natural oligomers of the amyloid-beta protein specifically disrupt cognitive function. *Nat Neurosci*. 2005 Jan;8(1): 79–84. <http://dx.doi.org/10.1038/nn1372>
112. Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, Wolfe MS, et al. Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature*. 2002 Apr 4;416(6880):535–9. <http://dx.doi.org/10.1038/416535a>

113. Lambert MP, Barlow AK, Chromy BA, Edwards C, Freed R, Liosatos M, et al. Diffusible, nonfibrillar ligands derived from A β 1-42 are potent central nervous system neurotoxins. *Proc Natl Acad Sci U S A*. 1998 May 26;95(11):6448–53. <http://dx.doi.org/10.1073/pnas.95.11.6448>
114. Lacor PN. Synaptic targeting by Alzheimer's-related amyloid oligomers. *J Neurosci*. 2004 Nov 10;24(45):10191–200. <http://dx.doi.org/10.1523/JNEUROSCI.3432-04.2004>
115. Shankar GM, Bloodgood BL, Townsend M, Walsh DM, Selkoe DJ, Sabatini BL. Natural oligomers of the Alzheimer amyloid-beta protein induce reversible synapse loss by modulating an NMDA-type glutamate receptor-dependent signaling pathway. *J Neurosci*. 2007 Mar 14;27(11):2866–75. <http://dx.doi.org/10.1523/JNEUROSCI.4970-06.2007>
116. Kent SA, Spiers-Jones TL, Durrant CS. The physiological roles of tau and A β : Implications for Alzheimer's disease pathology and therapeutics. *Acta Neuropathol*. 2020 Jul 29;140:417–47. <http://dx.doi.org/10.1007/s00401-020-02196-w>
117. Iqbal K, Liu F, Gong C-X, Grundke-Iqbal I. Tau in Alzheimer disease and related tauopathies. *Curr Alzheimer Res*. 2010 Dec;7(8):656–64. <http://dx.doi.org/10.2174/156720510793611592>
118. Gao Y, Tan L, Yu J-T, Tan L. Tau in Alzheimer's disease: Mechanisms and therapeutic strategies. *Curr Alzheimer Res*. 2018 Jan 23;15(3):1005–13. <http://dx.doi.org/10.2174/1567205014666170417111859>
119. Chen S, Townsend K, Goldberg TE, Davies P, Conejero-Goldberg C. MAPT isoforms: Differential transcriptional profiles related to 3R and 4R splice variants. *J Alzheimer's Dis*. 2011 Jan 7;22(4):1313–29. <http://dx.doi.org/10.3233/JAD-2010-101155>
120. Combs B, Mueller RL, Morfini G, Brady ST, Kanaan NM. Tau and axonal transport misregulation in tauopathies. *Adv Exp Med Biol*. 2019;1184(3):81–95. http://dx.doi.org/10.1007/978-981-32-9358-8_7
121. Yoshiyama Y, Higuchi M, Zhang B, Huang S-M, Iwata N, Saido TC, et al. Synapse loss and microglial activation precede tangles in a P301S tauopathy mouse model. *Neuron*. 2007 Feb;53(3):337–51. <http://dx.doi.org/10.1016/j.neuron.2007.01.010>
122. Allen B, Ingram E, Takao M, Smith MJ, Jakes R, Virdee K, et al. Abundant tau filaments and nonapoptotic neurodegeneration in transgenic mice expressing human P301S tau protein. *J Neurosci*. 2002 Nov 1;22(21):9340–51. <http://dx.doi.org/10.1523/JNEUROSCI.22-21-09340.2002>
123. Noble W, Pooler AM, Hanger DP. Advances in tau-based drug discovery. *Expert Opin Drug Discov*. 2011 Aug 24;6(8):797–810. <http://dx.doi.org/10.1517/17460441.2011.586690>
124. Brunden KR, Trojanowski JQ, Lee VM-Y. Advances in tau-focused drug discovery for Alzheimer's disease and related tauopathies. *Nat Rev Drug Discov*. 2009 Oct;8(10):783–93. <http://dx.doi.org/10.1038/nrd2959>
125. Stamer K, Vogel R, Thies E, Mandelkow E, Mandelkow E-M. Tau blocks traffic of organelles, neurofilaments, and APP vesicles in neurons and enhances oxidative stress. *J Cell Biol*. 2002 Mar 18;156(6):1051–63. <http://dx.doi.org/10.1083/jcb.200108057>
126. Dixit R, Ross JL, Goldman YE, Holzbaur ELF. Differential regulation of dynein and kinesin motor proteins by tau. *Science*. 2008 Feb 22;319(5866):1086–9. <http://dx.doi.org/10.1126/science.1152993>
127. Hoover BR, Reed MN, Su J, Penrod RD, Kotilinek LA, Grant MK, et al. Tau mislocalization to dendritic spines mediates synaptic dysfunction independently of neurodegeneration. *Neuron*. 2010 Dec;68(6):1067–81. <http://dx.doi.org/10.1016/j.neuron.2010.11.030>
128. Miller EC, Teravskis PJ, Dummer BW, Zhao X, Haganir RL, Liao D. Tau phosphorylation and tau mislocalization mediate soluble A β oligomer-induced AMPA glutamate receptor signaling deficits. *Eur J Neurosci*. 2014 Apr;39(7):1214–24. <http://dx.doi.org/10.1111/ejn.12507>
129. Wang Y, Mandelkow E. Tau in physiology and pathology. *Nat Rev Neurosci*. 2016;17(1):5–21. <http://dx.doi.org/10.1038/nrn.2015.1>
130. Shipton OA, Leitz JR, Dworzak J, Acton CEJ, Tunbridge EM, Denk F, et al. Tau protein is required for amyloid β -induced impairment of hippocampal long-term potentiation. *J Neurosci*. 2011;31(5):1688–92. <http://dx.doi.org/10.1523/JNEUROSCI.2610-10.2011>
131. Pickett EK, Herrmann AG, McQueen J, Abt K, Dando O, Tulloch J, et al. Amyloid beta and tau cooperate to cause reversible behavioral and transcriptional deficits in a model of Alzheimer's disease. *Cell Rep*. 2019 Dec;29(11):3592–604.e5. <http://dx.doi.org/10.1016/j.celrep.2019.11.044>

132. Spires-Jones TL, Hyman BT. The intersection of amyloid beta and tau at synapses in Alzheimer's disease. *Neuron*. 2014 May;82(4):756–71. <http://dx.doi.org/10.1016/j.neuron.2014.05.004>
133. Swerdlow RH, Burns JM, Khan SM. The Alzheimer's disease mitochondrial cascade hypothesis. Zhu X, Beal MF, Wang X, Perry G, Smith MA, editors. *J Alzheimer's Dis*. 2010 Jun 3;20(s2):S265–79. <http://dx.doi.org/10.3233/JAD-2010-100339>
134. Cenini G, Voos W. Mitochondria as potential targets in Alzheimer disease therapy: An update. *Front Pharmacol*. 2019;10(July):1–20. <http://dx.doi.org/10.3389/fphar.2019.00902>
135. Vetrivel KS, Thinakaran G. Membrane rafts in Alzheimer's disease beta-amyloid production. *Biochim Biophys Acta*. 2010 Aug;1801(8):860–7. <http://dx.doi.org/10.1016/j.bbailp.2010.03.007>
136. Swerdlow RH. Pathogenesis of Alzheimer's disease. *Clin Interv Aging*. 2007;2(3):347–59.
137. Hauptmann S, Scherping I, Dröse S, Brandt U, Schulz KL, Jendrach M, et al. Mitochondrial dysfunction: An early event in Alzheimer pathology accumulates with age in AD transgenic mice. *Neurobiol Aging*. 2009 Oct;30(10):1574–86. <http://dx.doi.org/10.1016/j.neurobiolaging.2007.12.005>
138. Du H, Guo L, Yan S, Sosunov AA, McKhann GM, Yan SS. Early deficits in synaptic mitochondria in an Alzheimer's disease mouse model. *Proc Natl Acad Sci U S A*. 2010;107(43):18670–5. <http://dx.doi.org/10.1073/pnas.1006586107>
139. Tamagno E, Guglielmotto M, Aragno M, Borghi R, Autelli R, Giliberto L, et al. Oxidative stress activates a positive feedback between the gamma- and beta-secretase cleavages of the beta-amyloid precursor protein. *J Neurochem*. 2008 Feb;104(3):683–95. <http://dx.doi.org/10.3389/fnins.2018.00830>
140. Ferreira LSS, Fernandes CS, Vieira MNN, De Felice FG. Insulin resistance in Alzheimer's disease. *Front Neurosci*. 2018;12(Nov):1–11. <http://dx.doi.org/10.3389/fnins.2018.00830>
141. Smythe GA, Bradshaw JE, Nicholson MV, Grunstein HS, Storlien LH. Rapid bidirectional effects of insulin on hypothalamic noradrenergic and serotonergic neuronal activity in the rat: Role in glucose homeostasis. *Endocrinology*. 1985;117(4):1590–7. <http://dx.doi.org/10.1210/endo-117-4-1590>
142. Shuaib A, Ijaz MS, Waqar T, Voll C, Kanthan R, Miyashita H, et al. Insulin elevates hippocampal GABA levels during ischemia. This is independent of its hypoglycemic effect. *Neuroscience*. 1995;67(4):809–14. [http://dx.doi.org/10.1016/0306-4522\(95\)00093-X](http://dx.doi.org/10.1016/0306-4522(95)00093-X)
143. Velliquette RA, O'Connor T, Vassar R. Energy inhibition elevates beta-secretase levels and activity and is potentially amyloidogenic in APP transgenic mice: Possible early events in Alzheimer's disease pathogenesis. *J Neurosci*. 2005 Nov 23;25(47):10874–83. <http://dx.doi.org/10.1523/JNEUROSCI.2350-05.2005>
144. Goyal MS, Vlassenko AG, Blazey TM, Su Y, Couture LE, Durbin TJ, et al. Loss of brain aerobic glycolysis in normal human aging. *Cell Metab*. 2017 Aug;26(2):353–60.e3. <http://dx.doi.org/10.1016/j.cmet.2017.07.010>
145. Mosconi L. Glucose metabolism in normal aging and Alzheimer's disease: Methodological and physiological considerations for PET studies. *Clin Transl Imaging*. 2013 Aug;1(4):1–7. <http://dx.doi.org/10.1007/s40336-013-0026-y>
146. Shah K, DeSilva S, Abbruscato T. The role of glucose transporters in brain disease: Diabetes and Alzheimer's disease. *Int J Mol Sci*. 2012;13(10):12629–55. <http://dx.doi.org/10.3390/ijms131012629>
147. Ding F, Yao J, Rettberg JR, Chen S, Brinton RD. Early decline in glucose transport and metabolism precedes shift to ketogenic system in female aging and Alzheimer's mouse brain: Implication for bioenergetic intervention. *PLoS One*. 2013;8(11):1–14. <http://dx.doi.org/10.1371/journal.pone.0079977>
148. Lee KY, Yoo DY, Jung HY, Baek L, Lee H, Kwon HJ, et al. Decrease in glucose transporter 1 levels and translocation of glucose transporter 3 in the dentate gyrus of C57BL/6 mice and gerbils with aging. *Lab Anim Res*. 2018;34(2):58. <http://dx.doi.org/10.5625/lar.2018.34.2.58>
149. Suh SW, Aoyama K, Chen Y, Garnier P, Matsumori Y, Gum E, et al. Hypoglycemic neuronal death and cognitive impairment are prevented by poly(ADP-ribose) polymerase inhibitors administered after hypoglycemia. *J Neurosci*. 2003;23(33):10681–90. <http://dx.doi.org/10.1523/JNEUROSCI.23-33-10681.2003>
150. Cheng B, Mattson MP. Glucose deprivation elicits neurofibrillary tangle-like antigenic changes in hippocampal neurons: Prevention by NGF and bFGF. *Exp Neurol*. 1992 Aug;117(2):114–23. [http://dx.doi.org/10.1016/0014-4886\(92\)90120-F](http://dx.doi.org/10.1016/0014-4886(92)90120-F)

151. Wang J, Alexanian A, Ying R, Kizhakekuttu TJ, Dharmashankar K, Vasquez-Vivar J, et al. Acute exposure to low glucose rapidly induces endothelial dysfunction and mitochondrial oxidative stress. *Arterioscler Thromb Vasc Biol.* 2012 Mar;32(3):712–20. <http://dx.doi.org/10.1161/ATVBAHA.111.227389>
152. Durrant CS, Ruscher K, Sheppard O, Coleman MP, Özen I. Beta secretase 1-dependent amyloid precursor protein processing promotes excessive vascular sprouting through NOTCH3 signalling. *Cell Death Dis.* 2020;11(2):98. <http://dx.doi.org/10.1038/s41419-020-2288-4>
153. Roher AE, Debbins JP, Malek-Ahmadi M, Chen K, Pipe JG, Maze S, et al. Cerebral blood flow in Alzheimer's disease. *Vasc Health Risk Manag.* 2012;8:599–611. <http://dx.doi.org/10.2147/VHRM.S34874>
154. Navarro-Dorado J, Villalba N, Prieto D, Brera B, Martín-Moreno AM, Tejerina T, et al. Vascular dysfunction in a transgenic model of Alzheimer's disease: Effects of CB1R and CB2R cannabinoid agonists. *Front Neurosci.* 2016;10(Sep):422. <http://dx.doi.org/10.3389/fnins.2016.00422>
155. Lee JW, Lee YK, Yuk DY, Choi DY, Ban SB, Oh KW, et al. Neuro-inflammation induced by lipopolysaccharide causes cognitive impairment through enhancement of beta-amyloid generation. *J Neuroinflammation.* 2008;5:1–14. <http://dx.doi.org/10.1186/1742-2094-5-37>
156. Sheppard O, Coleman MP, Durrant CS. Lipopolysaccharide-induced neuroinflammation induces pre-synaptic disruption through a direct action on brain tissue involving microglia-derived interleukin 1 beta. *J Neuroinflammation.* 2019 May 18;16(1):106. <http://dx.doi.org/10.1186/s12974-019-1490-8>
157. Chen YY, Zhang L, Shi DL, Song XH, Shen YL, Zheng MZ, et al. Resveratrol attenuates subacute systemic inflammation-induced spatial memory impairment via inhibition of astrocyte activation and enhancement of synaptophysin expression in the hippocampus. *Ann Clin Lab Sci.* 2017;47(1):17–24.
158. Rao JS, Kellom M, Kim H-W, Rapoport SI, Reese EA. Neuroinflammation and synaptic loss. *Neurochem Res.* 2012 May;37(5):903–10. <http://dx.doi.org/10.1007/s11064-012-0708-2>
159. Castaño A, Herrera AJ, Cano J, Machado A. Lipopolysaccharide intranigral injection induces inflammatory reaction and damage in nigrostriatal dopaminergic system. *J Neurochem.* 1998 Apr 14;70(4):1584–92. <http://dx.doi.org/10.1046/j.1471-4159.1998.70041584.x>
160. Sheng JG, Bora SH, Xu G, Borchelt DR, Price DL, Koliatsos VE. Lipopolysaccharide-induced-neuroinflammation increases intracellular accumulation of amyloid precursor protein and amyloid β peptide in APP^{Swe} transgenic mice. *Neurobiol Dis.* 2003;14(1):133–45. [http://dx.doi.org/10.1016/S0969-9961\(03\)00069-X](http://dx.doi.org/10.1016/S0969-9961(03)00069-X)
161. Wang L-M, Wu Q, Kirk RA, Horn KP, Ebada Salem AH, Hoffman JM, et al. Lipopolysaccharide endotoxemia induces amyloid- β and p-tau formation in the rat brain. *Am J Nucl Med Mol Imaging.* 2018 Nov;8(2):86–99.
162. Neniskyte U, Neher JJ, Brown GC. Neuronal death induced by nanomolar amyloid β is mediated by primary phagocytosis of neurons by microglia. *J Biol Chem.* 2011;286(46):39904–13. <http://dx.doi.org/10.1074/jbc.M111.267583>
163. Stalder M, Phinney A, Probst A, Sommer B, Staufenbiel M, Jucker M. Association of microglia with amyloid plaques in brains of APP23 transgenic mice. *Am J Pathol.* 1999;154(6):1673–84. [http://dx.doi.org/10.1016/S0002-9440\(10\)65423-5](http://dx.doi.org/10.1016/S0002-9440(10)65423-5)
164. Bolmont T, Haiss F, Eicke D, Radde R, Mathis CA, Klunk WE, et al. Dynamics of the microglial/amyloid interaction indicate a role in plaque maintenance. *J Neurosci.* 2008;28(16):4283–92. <http://dx.doi.org/10.1523/JNEUROSCI.4814-07.2008>
165. Yang W, Tao Y, Wu Y, Zhao X, Ye W, Zhao D, et al. Neutrophils promote the development of reparative macrophages mediated by ROS to orchestrate liver repair. *Nat Commun.* 2019 Dec 6;10(1):1076. <http://dx.doi.org/10.1038/s41467-019-09046-8>
166. Flohé L, Brigelius-Flohé R, Saliou C, Traber MG, Packer L. Redox regulation of NF-kappa B activation. *Free Radic Biol Med.* 1997;22(6):1115–26. [http://dx.doi.org/10.1016/S0891-5849\(96\)00501-1](http://dx.doi.org/10.1016/S0891-5849(96)00501-1)
167. Anderson MT, Staal FJT, Gitler C, Herzenberg LA, Herzenberg LA. Separation of oxidant-initiated and redox-regulated steps in the NF- κ B signal transduction pathway. *Proc Natl Acad Sci U S A.* 1994;91(24):11527–31. <http://dx.doi.org/10.1073/pnas.91.24.11527>
168. Tan BL, Norhaizan ME, Liew WPP, Rahman HS. Antioxidant and oxidative stress: A mutual interplay in age-related diseases. *Front Pharmacol.* 2018;9(Oct):1–28. <http://dx.doi.org/10.3389/fphar.2018.01162>

169. Bemiller SM, McCray TJ, Allan K, Formica SV, Xu G, Wilson G, et al. TREM2 deficiency exacerbates tau pathology through dysregulated kinase signaling in a mouse model of tauopathy. *Mol Neurodegener.* 2017;12(1):1–12. <http://dx.doi.org/10.1186/s13024-017-0216-6>
170. Ising C, Venegas C, Zhang S, Scheiblich H, Schmidt SV, Vieira-Saecker A, et al. NLRP3 inflammatory activation drives tau pathology. *Nature.* 2019;575(7784):669–73. <http://dx.doi.org/10.1038/s41586-019-1769-z>
171. Morales I, Jiménez JM, Mancilla M, Maccioni RB. Tau oligomers and fibrils induce activation of microglial cells. *J Alzheimer's Dis.* 2013;37(4):849–56. <http://dx.doi.org/10.3233/JAD-131843>
172. Lastres-Becker I, Innamorato NG, Jaworski T, Rábano A, Kügler S, Van Leuven F, et al. Fractalkine activates NRF2/NFE2L2 and heme oxygenase 1 to restrain tauopathy-induced microgliosis. *Brain.* 2014;137(1):78–91. <http://dx.doi.org/10.1093/brain/awt323>
173. Vogels T, Murgoci A-N, Hromádka T. Intersection of pathological tau and microglia at the synapse. *Acta Neuropathol Commun.* 2019 Nov 5;7(1):109. <http://dx.doi.org/10.1186/s40478-019-0754-y>
174. Jay TR, Miller CM, Cheng PJ, Graham LC, Bemiller S, Broihier ML, et al. TREM2 deficiency eliminates TREM2+ inflammatory macrophages and ameliorates pathology in Alzheimer's disease mouse models. *J Exp Med.* 2015;212(3):287–95. <http://dx.doi.org/10.1084/jem.20142322>
175. Thibautaud TA, Anderson RT, Smith DM. A common mechanism of proteasome impairment by neurodegenerative disease-associated oligomers. *Nat Commun.* 2018;9(1):1097. <http://dx.doi.org/10.1038/s41467-018-03509-0>
176. Upadhyya SC, Hegde AN. Role of the ubiquitin proteasome system in Alzheimer's disease. *BMC Biochem.* 2007 Nov 22;8(Suppl. 1):S12. <http://dx.doi.org/10.1186/1471-2091-8-S1-S12>
177. Oddo S. The ubiquitin-proteasome system in Alzheimer's disease. *J Cell Mol Med.* 2008;12(2):363–73. <http://dx.doi.org/10.1111/j.1582-4934.2008.00276.x>
178. Liu J, Li L. Targeting autophagy for the treatment of Alzheimer's disease: Challenges and opportunities. *Front Mol Neurosci.* 2019;12(August):1–9. <http://dx.doi.org/10.3389/fnmol.2019.00203>
179. Haass C, Kaether C, Thinakaran G, Sisodia S. Trafficking and proteolytic processing of APP. *Cold Spring Harb Perspect Med.* 2012 May 1;2(5):a006270. <http://dx.doi.org/10.1101/cshperspect.a006270>
180. Yin RH, Yu JT, Tan L. The role of SORL1 in Alzheimer's disease. *Mol Neurobiol.* 2015;51(3):909–18. <http://dx.doi.org/10.1007/s12035-014-8742-5>
181. Glennon EBC, Whitehouse IJ, Miners JS, Kehoe PG, Love S, Kellett KAB, et al. BIN1 is decreased in sporadic but not familial Alzheimer's disease or in aging. *PLoS One.* 2013;8(10):1–11. <http://dx.doi.org/10.1371/journal.pone.0078806>
182. Francis PT, Palmer AM, Snape M, Wilcock GK. The cholinergic hypothesis of Alzheimer's disease: A review of progress. *J Neurol Neurosurg Psychiatry.* 1999;66(2):137–47. <http://dx.doi.org/10.1136/jnnp.66.2.137>
183. Craig LA, Hong NS, McDonald RJ. Revisiting the cholinergic hypothesis in the development of Alzheimer's disease. *Neurosci Biobehav Rev.* 2011 May;35(6):1397–409. <http://dx.doi.org/10.1016/j.neubiorev.2011.03.001>
184. Hardy J. A hundred years of Alzheimer's disease research. *Neuron.* 2006 Oct 5;52(1):3–13. <http://dx.doi.org/10.1016/j.neuron.2006.09.016>

